ANNALES UNIVERSITATIS MARIAE CURIE-SKŁODOWSKA LUBLIN-POLONIA

VOL. LIII, 15

SECTIO DD

1998

Katedra Patofizjologii Wydziału Medycyny Wetrynaryjnej AR oraz Katedra Chorób Wewnętrznych AM w Lublinie

ANDRZEJ LEDWOŻYW, MONIKA BOJARSKA - ŁOŚ

Determination of plasminogen/plasmin levels in horse, cattle, sheep and pig plasma by the use of synthetic chromogenic substrate

Oznaczanie stężenia plazminogenu/plazminy w osoczu krwi koni, bydła, owiec i świń przy użyciu syntetycznego substratu chromonogennego

The fibrinolytic system comprises a proenzyme, plasminogen, which can be converted to the active enzyme, plasmin, which degrades fibrin. Plasminogen activation is mediated by plasminogen activators, which are classified as either tissue-type plasminogen activators (t-PA) or urokinase-type plasminogen activators (u-PA). Inhibition of the fibrinolytic system may occur at the level of the activators or at the level of activated plasmin.

Plasmin has a low substrate specificity, and when circulating freely in the blood it degrades several proteins including fibrinogen, factor V, and factor VIII. Plasma does, however, contain a fast-acting plasmin inhibitor, α_2 -antiplasmin, which inhibits free plasmin extremely rapidly but which reacts much slower with plasmin bound to fibrin (7).

Plasminogen activation is the main regulatory process in fibrinolytic system. Fibrinolysis may occur by three different ways: a) by t-PA-dependent way, b) by factor XII-dependent way and c) by factor XII-independent way (17).

t-PA is secreted by endothelial cells and transported to blood circulation. The concentration of this Protein is 5 - 50 ng/ml (33) and biological half-life 2.5 min. (3). At this level plasminogen activator inhibitor (PAI) is active (37); liver or vascular endothelium may be the sources of PAI. Units of t-AP activity are based on urokinase units (15). The level of t-AP in blood circulation in man is 0.1 - 0.4 IU/ml and the level of PAI is 5 - 10 IU/ml (38). This, in normal conditions an excess of PAI exists. Diurnal changes in fibrinolytic activity are caused by the changes in PAI activity (21). The decreased t-AP activity in early postoperative period and after heart infarct is also caused by PAI activity increase (12). The rapid changes observed in these states are caused by short half-life of PAI (11). Factor XII-dependent plasminogen activation is one of the most intriguing haemostasis processes. Factor XII after activation to factor XIIa cause the change of Precallicrein to callicrein and the last one is a strong plasminogen activator (10).

Factor XII-independent way is urokinase-dependent (3). Leukocyte-derived plasminogen activators enhanced excretion is the cause of coagulation disorders in Chediak-Higashi syndrome (13).

Plasmin is a key enzyme in fibrinolytic system. The enzyme shows a broad, trypsin-like specificity and its unspecific proteolytic activity is regulated in blood circulation by protease inhibitors, such as α_2 -antiplasmin. Its inheritant deficiency or absence cause bleeding tendency

(19, 20). Both plasminogen and -antiplasmin are modulated through many factors. In normal conditions, only 60% of plasminogen is available for activation; the remaining portion of this protein is connected with circulating histidine-rich glycoprotein (HRG) (28). Another plasminogen-binding protein, tetranectin, also exists (6).

Thrombolytic activity of t-AP was examined on animal experimental models of lung infarct (4, 30), venous thrombosis (8, 9) and coronary artery thrombosis (14, 16).

The synthesis of synthetic substrate, sensitive to plasmin activity, was essential in the development of the methods of precise plasmin gen/plasmin level estimation in human blood plasma (5, 18).

In previous publications (24, 25, 26, 27) we have stated the usefulness of chromogenic substrates for the estimation of platelet factor 3, endogenous heparin and factor VII and VIII levels in blood plasma of domestic animals.

The aim of this study was to evaluate the suitability of the synthetic tripeptide substrate, H-D-Valyl-Leucyl-Lysyl-p-nitroanilide (S-2251) in the estimation of plasminogen/plasmin concentration in blood plasma of domestic animals.

MATERIAL AND METHODS

Principle of the procedure

Plasminogen + Streptokinase (excess) -----> Plasminogen-streptokinase complex H-D-Val-Leu-Lys-p-NA Plasminogen-streptokinase complex H-D-Val-Leu-Lys + p-NA

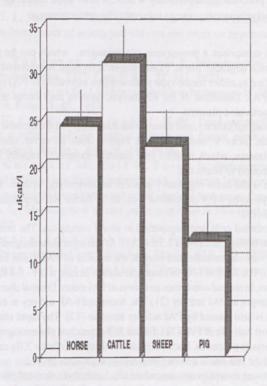


Fig. 1. Plasminogen/plasmin levels in horse (n = 21), cattle (n = 20), sheep (n = 19) and pig (n = 24) plasma. Mean \pm S.D.

Plasminogen is converted into an active plasminogen-streptokinase complex when an excess of streptokinase is added. The complex catalyzes splitting of p-nitroaniline (p-NA) from the substrate. The rate at which p-NA is released is measured photometrically at 405 nm.

The blood of 21 horses, 20 cows, 19 sheep and 24 pigs (9 vol) was mixed with 1 vol. of 0.1 M sodium citrate solution and centrifuged at 2000 x g for 20 min.

To standard curve construction, a human plasmin according to 1st British Standard for Plasmin (National Institute for Biological Standards and Controls, London, England) was used for no availability of animal plasmin specimen or standards. The 2 mM substrate (S-2251) solution and streptokinase (4 000 IU/ml) were purchased from Kabi - Vitrum (Molndal, Sweden).

Before estimations, $50 \,\mu l$ plasma was mixed with 2 ml of Tris-HCl buffer (0.05 M, pH = 7.4).

To thermostated (37 $^{\circ}$ C) spectrophotometer cuvette (Specord UV-VIS, Karl Zeiss, Jena, Germany) 200 μ l of diluted plasma was poured, and after 4-5 min 200 μ l of streptokinase solution was added. Mixture was incubated for 10 min. After incubation period 200 of S-2251 was added and E₄₀₅/min was measured.

One katal (kat) is defined as the amount of enzyme activity that will release one mole of P-nitroaniline per second.

RESULTS

Figure 1 shows the plasmin activity in blood plasma of examined animal species. The highest activity of plasmin was stated in cattle, the lowest one in pig.

Figure 2 shows the velocity of p-nitroaniline release from S-2251 substrate as dependent on the time of reaction and plasmin concentration. By the incubations

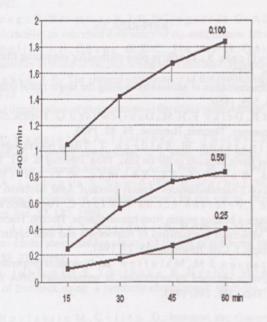


Fig. 2. The rate of p-nitroaniline release from chromogenic substrate as dependent on incubation time (horse plasma, n = 21). Mean \pm S.D.

time above 10 min. a dependence was observed. To keep the proportionality between hydrolysis rate and enzyme concentration, the time of incubation of 10 min was chosen.

DISCUSSION

The elevated plasminogen levels were stated in acute bacterial infections, inflammatory states, thrombophlebitis, after major surgery, in myocardial infarction, in pregnancy, the lowered one in disseminated intravascular coagulation (DIC) and in liver cirrhosis (22). The plasminogen level is routinely measured by the use of caseinolytic (18, 34) and fibrinolytic (1, 23) methods, immunological methods are also available (2, 29). However, the long incubation time in radial immunodiffusion methods (a dozen or so hours) keep away the rapid plasminogen level estimations. The caseinolytic method is both laborious and time-consuming.

The amidolytic methods with the use of chromogenic substrates based on the observation that streptokinase forms an equimolar complex with plasminogen (32, 35). Introduction of synthetic substrates (5, 31, 36) allows to elaborate sensitive and rapid methods, useable in laboratory practice.

The results presented above suggest that the S-2251 substrate is entirely useful in plasminogen activity determination in blood plasma of large domestic animals.

REFERENCES

- 1. A strup T., Mullertz S.: The fibrin plate method for estimating fibrinolytic activity. Arch. Biochem. 40, 346, 1952.
- Becker W.: Determination of antisera titres using the single radial immunodiffusion method. Immunochem. 6, 539, 1969.
- 3. Brommer E. J. P., Derk x F. H. M., Dooijewaard G.: Hepatic clearance of endogenous fibrinolytic components. Thromb. Haemost. 54, 42, 1985.
- 4. Carlin G., Finarsson M., Saldeen T.: Tissue plasminogen activator effectively lyses intravascular fibrin deposits in the rat lung. Prog. Fibrinol. 6, 471, 1983.
- 5. Clavin S. A., Bobbitt J. L., Shuman R. T., Smithwick E. L.: Use of peptidyl-4-methoxy-2-naphthylamides to assay plasmin. Anal. Biochem. 80, 355, 1977.
- 6. Clemmensen I., Petersen L., Los P., Kluft C.: Purification and characterization of a plasminogen kringle 4 binding protein from human plasma. Thromb. Haemost. 54, 277, 1985.
- C o I I e n D.: Molecular mechanisms of fibrinolysis and their application to fibrin-specific thrombolytic therapy. J. Cell. Biochem. 33, 77, 1987.
- 8. Collen D., Stassen J. M., Marafino B.: Biological properties of human tissue-type plasminogen activator obtained by expression of recombinant DNA in mammalian cells. J. Pharmacol. Exp. Ther. 231, 146, 1983.

- 9. Collen D., Stassen J. M., Verstraete M.: Thrombolysis with human extrinsic (tissue-type) plasminogen activator in rabbits with experimental jugular vein thrombosis. Effect of molecular form and dose of activator, age of the thrombus, and route of administration. J. Clin. Invest. 73, 368, 1983.
- 10. Colman R. W.: Activation of plasminogen by human plasma kallikrein. Biochem. Biophys. Res. Commun. 35, 273, 1969.
- 11. Colucci M., Parano J. A., Collen D.: Generation in plasma of a fast-acting inhibitor of plasminogen activator in response to endotoxin stimulation. J. Clin. Invest. 75, 818, 1985.
- Dangelo A., Kluft C., Verheijen J. H.: Fibrinolytic shunt-down after surgery: impairment of the balance between tissue-type plasminogen activator and its specific inhibitor. Eur. J. Clin. Invest. 15, 308, 1985.
- 13. De Saint Basile G., Fischer A., Dautzenberg M. D.: Enhanced plasminogen activator production by leukocytes in the human and murine Chediak-Higashi syndrome. Blood 65, 1275, 1985.
- 14. Flameng V., Van De Verf F., Vanhaecke J.: Coronary thrombolysis and infarct size reduction following intravenous infusion of recombinant tissue-type plasminogen activator in non-human primates. J. Clin. Invest. 75, 84, 1985.
- 15. Gaffney P. J., Curtis A. D.: A collaborative study of a proposed international standard for tissue plasminogen activator (t-PA). Thromb. Haemost. 53, 134, 1985.
- 16. Gold H. K., Fallon J. T., Yasuda T.: Coronary thrombolysis with recombinant human tissue plasminogen activator. Circulation 70, 700, 1984.
- 17. Hessel L. W., Kluft C.: Advances in clinical fibrinolysis. Clin. Haematol. 15, 443, 1986.
- Johnson A. J., Kline D. L., Alkjaersig N.: Assay methods and standard preparations for plasmin, plasminogen and urokinase in purified systems. Thromb. Diath. Haemorrh. 21, 259, 1969.
- Kettle P., Mayne E. E.: A bleeding disorder due to deficiency of α₂-antiplasmin. J. Clin. Pathol. 38, 428, 1985.
- Kluft C., Vellenga E., Brommer K. J. P., Wijngaards G.: A familial haemorrhagic diathesis in a Dutch family: an inherited deficiency of α₂-antiplasmin. Blood 59, 1169, 1982.
- 21. Kluft C., Verheijen J. H., Chang G. T. C., Jie A. F. H., Onkelinx C.: Diurnal fluctuations in the activity in blood of the fast-acting t-PA inhibitor. Haemostasis. 100, 182, 1984.
- 22. Lackner H., Javid J. P.: The clinical significance of the plasminogen level. Am. J. Clin. Pathol. 60, 175, 1973.
- 23. Lassen M.: Heat denaturation of plasminogen in the fibrin plate method. Acta Physiol. Scand. 27, 371, 1952.
- 24. Ledwożyw A., Kądziołka A., Jabłonka S., Tusińska E., Herbut M.: Oznaczanie aktywności czynnika płytkowego 3 u koni, bydła, owiec i świń przy użyciu chromogennego substratu trójpeptydowego. Pol. Arch. Wet. 31, 71, 1991.
- 25. Ledwożyw A., Jabłonka S., Tusińska E., Herbut M.: The estimation of factor VII in livestock plasma of domestic animals by the use of tripeptide chromogenic substrate. Arch. Vet. Pol. 33, 123, 1993.
- 26. Ledwożyw A., Jabłonka S., Tusińska E., Herbut M.: The estimation of factor VIII levels in horse, cattle, sheep and pig plasma by the use of synthetic chromogenic substrate. Arch. Vet. Polon. 33, 275, 1993.
- Ledwożyw A., Jabłonka S., Tusińska E., Herbut M.: Assay for endogenous heparin in plasma of livestock using a synthetic chromogenic substrate. Arch. Vet. Polon. 33, 269, 1993.
- 28. Lijnen H. R., Hoylaerts M., Collen D.: Isolation and characterization of a human plasma protein with affinity for the lysine binding sites in plasminogen. Role in the regulation of fibrinolysis and identification as histidine-rich glycoprotein. J. Biol. Chem. 255, 10214, 1980.

- 29. Mancini G., Carbonera A. O., Hevemans J. F.: Immunochemical quantitation of antigens by single radial immunodiffusion. Immunochem. 2, 235, 1965.
- 30. Matsuo O., Rijken D. C., Collen D.: Thrombolysis by human tissue plasminogen activator and urokinase in rabbits with experimental pulmonary embolism. Nature 291, 590, 1981.
- 31. Mattler L. E., Bang N. U.: Serine protease specificity for peptide chromogenic substrates. Thromb. Haemost. 38, 776, 1977.
- 32. McClintock D. K., Bell P. H.: The mechanism of activation of human plasminogen by streptokinase. Biochem. Biophys. Res. Commun. 43, 694, 1971.
- 33. Nilsson I. M., Ljungner H., Tengborn L.: Two different mechanisms in patient with venous thrombosis and defective fibrinolysis: low concentrations of plasminogen activator or increased concentration of plasminogen activator inhibitor. Br. Med. J. 290, 1453, 1985.
- 34. Rennert L. F., Cohen P. P.: Partial purification and properties of a proteolytic enzyme of human serum. J. Biol. Chem. 181, 431, 1949.
- 35. Schick L. A., Castellino F. J.: Direct evidence for the generation of an active-site in the plasminogen moiety of the streptokinase-human plasminogen activator complex. Biochem. Biophys. Res. Commun. 57, 47, 1974.
- 36. Teger-Nilsson A. C., Friberger P., Gyzander E.: Determination of a new rapid plasmin inhibitor in human blood by means of a plasmin-specific tripeptide substrate. Scand. J. Clin. Lab. Invest. 37, 403, 1977.
- Verheijen J. H., Chang G. T. G., Kluft C.: Evidence for the occurrence of a fast-acting inhibitor for tissue-type plasminogen activator in human plasma. Thromb. Haemost. 51, 392, 1984
- 38. Wiman B., Ljungberg B., Chmielewska J.: The role of the fibrinolytic system in deep vein thrombosis. J. Lab. Clin. Med. 105, 265, 1985.
- 39. Wun T. C., Schleuning W. D., Reich E.: Isolation and characterization of urokinase from human plasma. J. Biol. Chem. 257, 3276, 1982.

STRESZCZNIE

Oznaczano stężenia plazminogenu/plazminy w osoczu krwi koni, bydła, owiec i świń przy użyciu chromogennego substratu trójpeptydowego p-nitroanilidu H-D-valilo-leucylo-lizyny (S-2251). Najwyższy poziom tego czynnika (31,2 mkat/l) stwierdzono w osoczu krwi bydła, najniższy zaś (12,3 mkat/l) u świń. W osoczu krwi koni i owiec poziomy te wynosiły odpowiednio 24,5 mkat/l i 22,3 mkat/l. Szybkość uwalniania p-nitroaniliny z substratu w zależności od czasu inkubacji w osoczu krwi koni wzrastała z 1,05 do 1,84 (ΔΕ405/min) przy 0,1 jednostek plazminy w objętości próbki, z 0,25 do 0,83 przy 0,5 jednostki plazminy w objętości próbki, z 0,1 do 0,4 przy 0,25 jednostki plazminy w objetości próbki

Stwierdzono przydatność substratu S-2251 do oznaczania stężenia plazminogenu/plazminy w osoczu krwi zwierząt gospodarskich.